# Flavonoids and total phenolic content in extruded buckwheat products with sweet and salty taste

G. Krumina–Zemture<sup>1,\*</sup>, I. Beitane<sup>1</sup> and I. Cinkmanis<sup>2</sup>

<sup>1</sup>Latvia University of Life Sciences and Technologies, Faculty of Food Technology, Department of Nutrition, Riga street 22, LV–3004, Jelgava, Latvia <sup>2</sup>Latvia University of Life Sciences and Technologies, Faculty of Food Technology, Department of Chemistry, Liela street 2, LV–3001, Jelgava, Latvia \*Correspondence: gita.krumina@llu.lv

Abstract. The aim of research was to evaluate the effect of added ingredients used for taste improvement on flavonoids and total phenolic content in extruded buckwheat products. The added ingredients were – sugar, vanilla sugar, stevia, agave syrup, cinnamon, caraway, garlic powder, sweet pepper powder and salt. Six extruded buckwheat products were analysed, where an extruded buckwheat product without added ingredients was a control sample, two extruded buckwheat products were with sweet taste and three products with salty taste. Total phenolic content was determined using the modified Folin–Ciocalteu method but flavonoid content according to LC-TOF-MS method.

The highest total phenolic content (p < 0.05) was determined in extruded buckwheat product with vanilla sugar + stevia (91.17  $\pm$  0.67 GAE mg 100g<sup>-1</sup>DW). It was almost three times higher than in the control sample. The significant differences were observed in extruded buckwheat products with caraway + salt + agave syrup (41.5  $\pm$  0.12 GAE mg 100g<sup>-1</sup>DW) and sweet pepper powder + salt + agave syrup (42.39  $\pm$  0.80 GAE mg 100g<sup>-1</sup>DW) comparing to other extruded products (p < 0.05). The highest content of rutin and quercetin (p < 0.05) was established in extruded buckwheat product with garlic powder + salt + agave syrup, whereas the highest content of catechin and epicatechin – in extruded buckwheat product with vanilla sugar + stevia. The extruded buckwheat product with caraway + salt + agave syrup in addition contained luteolin, kaempferol and isoquercitrin. The results of research showed that some added ingredients used for taste improvement can significantly influence the total phenolic content and flavonoid content.

Key words: buckwheat, extrusion, flavonoids, total phenolic content.

# **INTRODUCTION**

Buckwheat is an alternative crop with high consumption in countries like China, Japan and Taiwan due to its functional food properties and biological value – great concentration of bio–active compounds such as phenols and flavonoids (Lin et al., 2009; Inglett et al., 2010; Qin et al., 2010; Zhang et al., 2012; Vollmannova et al., 2013). The diverse total phenolic content of buckwheat is reported in literature, the differences of results establish buckwheat species: common (*Fagopyrum esculentum* Moench) or tartary (*Fagopyrum tataricum* Gaertn); cultivars, growth conditions and technological production process. Mikulajeva et al. (2016) analysed 22 common buckwheat cultivars,

which total phenolic content ranged between 0.897 and 4.226 mg GAE g<sup>-1</sup> DW. Unal et al. (2017) reported about significant differences of total phenolic content between commercial buckwheat (produced in Kazakhstan) and Günes variety (produced in Turkey) – 207.12 ± 2.67 and 329.83 ± 3.88 mg GAE 100 g<sup>-1</sup>, respectively. The diverse results about flavonoids content like rutin can be found in literature, too. Mikulajova et al. (2016) established rutin content in common buckwheat cultivars between  $59.93 \pm 2.16$  and  $304.45 \pm 5.45 \ \mu g \ g^{-1}$  DW, whereas Vollmannova et al. (2013) – between  $309.97 \pm 10.84$  and  $507.78 \pm 17.11 \ mg \ kg^{-1}$  DW.

The increase of buckwheat consumption in Latvia could be possible by production of new buckwheat products. In Asian countries and Italy there are buckwheat noodles, in Latvia there could be some buckwheat snacks using extrusion. There is not a single view about the effect of processing on phenolic and flavonoid content in buckwheat. Zielinska et al. (2007) and Wronkowska et al. (2015) concluded significant differences of phenolic compounds in raw and roasted buckwheat groats, whereas Hes et al. (2014) did not establish any negative effect on the nutritional properties in buckwheat groats after 30 min boiling in water. Sensoy et al. (2006) determined insignificant decrease of total phenolic content in dark buckwheat flour after roasting (200 °C, 10 min), whereas antioxidant activity was significantly lower in roasted dark buckwheat flour comparing to raw dark buckwheat flour. Our previous research (Beitane et al., 2018) showed significant decrease of total phenolic content, antiradical activity, radical scavenging activity and phenolic compounds concentration in buckwheat after extrusion.

In production of new products their taste has got important role, which could be improved with several ingredients. Furthermore the added ingredients used for taste improvement can affect the nutritional and biological value of the final product. In this research the added ingredients for taste improvement of extruded buckwheat products were sugar, vanilla sugar, stevia, agave syrup, cinnamon,-caraway, garlic powder, sweet pepper powder and salt. The brief description of used ingredients and their importance in food product production is given below.

Vanillin is the most used flavour in the food industry with antimicrobial and antioxidant properties (Zabkova et al., 2006).

Stevia rebaudiana or steviol glycosides are thermally stable and natural sweeteners with high sweetening potential, which are successfully used in cocoa and chocolate products, in chewing gums, flour or starch–based snacks etc. to reduce sugar consumption in the industrialized countries (Olsson et al., 2016; Shannon et al., 2016).

Agave syrup is a natural sweet product with functional properties due to biological active compounds, prebiotic molecules and antioxidant content, which is used as sugar replacement due to low glycaemic index (Foster–Powell et al., 2002; Phillips et al., 2009; Mellado–Mojica & Lopez, 2015, Muniz–Marquez et al., 2015).

Cinnamon is a culinary spice with anti-oxidant properties and free radical scavenging activity, which is used in traditional Chinese medicine and which has beneficial effects on serum glucose, triglyceride and total cholesterol level in diabetic patients (Parkash et al., 2007; Crawford, 2009; Otto, 2010; Adisakwattana et al., 2011).

Caraway is the most cultivated spice in Europe with wide use in folk medicine (Ghoneem et al., 2016; Kluz et al., 2016). Particular attention in scientific researches is paid to caraway essential oil with the main components – carvone and limonene (Ghoneem et al., 2016).

Garlic is a vegetable, which is known worldwide, with anti-bacterial and antifungal properties, high flavonoids content, especially quercetin, and antioxidant capacity (Griffiths et al., 2002; Beato et al., 2011; Onyeoziri et al., 2016).

Sweet pepper is a vegetable with high content of vitamins, carotenoids and phenols, which could be included in daily diet or used as additive in food production or in traditional medicine (Marti et al., 2011; Hernandez–Ortega et al., 2012; Sharma et al., 2016; Raybaudi–Massilia et al., 2017)

The aim of research was to evaluate the effect of added ingredients used for taste improvement on flavonoids and total phenolic content in extruded buckwheat products.

# **MATERIALS AND METHODS**

#### Materials

Raw common buckwheat (Fagopyrum esculentum Moench) grown in Latvia in 2017, was produced by the organic farm 'Bebri'.

The added ingredients for taste improvement of extruded buckwheat products were purchased in grocery: sugar (Dan Sukker, Denmark), vanilla sugar produced from sugar and flavouring material vanillin (Mood, Lithuania), stevia (Canderel, The Czech Republic), agave syrup (Super Garden, Mexico), cinnamon (Santa Maria, Estonia), caraway (Santa Maria, Estonia), garlic powder (Kotanyi, Austria), sweet pepper powder (Santa Maria, Estonia) and salt (Artiomosol, Ukraine).

### **Dough preparation for extrusion**

Raw buckwheat grain was milled using Mühle 2 (Hawo's, Germany) to obtain whole grain flour. Buckwheat flour with added ingredients and water was mixed with Sirman Planetary Mixers, model Pluton 10 (Minneapolis, Italy). The added ingredients for taste improvement of extruded buckwheat products were sugar, vanilla sugar, stevia, agave syrup, cinnamon, caraway, garlic powder, sweet pepper powder and salt. Six different doughs were prepared (Table 1), where buckwheat dough without added ingredients was prepared as control, two doughs were with sweet taste (vanilla sugar + stevia; sugar + cinnamon) and three – with salty taste (caraway + salt + agave syrup; garlic powder + salt + agave syrup; sweet pepper powder + salt + agave syrup).

	Control	B+VS	B+SC	B+CSA	B+GSA	B+PSA
Buckwheat flour, g	100	100	100	100	100	100
Water, mL	75	75	75	75	75	75
Sugar, g	_	_	5	_	_	_
Vanilla sugar, g	—	5	_	—	—	—
Stevia, g	—	4	_	—	—	—
Agave syrup, g	—	_	_	5	5	5
Cinnamon, g	—	_	3	—	—	—
Caraway, g (milled)	—	_	_	5	—	_
Garlic powder, g	—	_	_	—	1.5	—
Sweet pepper powder, g	—	_	_	—	—	5
Salt, g	_	<u> </u>	_	1.5	1.5	1.5

Table 1. The formation of dough and abbreviations in buckwheat products

#### Extrusion

The prepared buckwheat doughs with different tastes were extruded with the food extruder PCE Extrusiometer L–Serie (Göttfert, Germany) by temperature profile 75/90/100 °C. After extrusion the products were cut in squares of 1 cm, dried in the convective–rotary oven (SVEBA DAHLAN, Sweden) at the temperature 140  $\pm$  2 °C for 25 min and cooled to room temperature. Afterwards, six acquired extruded buckwheat products were used for further analysis.

#### Methods

Total phenolic content (TPC) of added ingredients for taste improvement and extruded buckwheat products was determined using the modified Folin–Ciocalteu method as described by Herald et al. (2012). The measurement was conducted by mixing Folin–Ciocalteu solution (1:1 with water), sodium bicarbonate and ethanolic extract. The absorbance was measured after 90 min of incubation at 765 nm againts a blank. TPC was expressed as gallic acid equivalents (GAE mg 100 g<sup>-1</sup> DW), based on the gallic acid (GA) calibration curve (range 0.025–0.20 mg mL<sup>-1</sup>, R<sup>2</sup> = 0.9997). Analyses were performed with the Infinite M200 PRO (Tecan Group Ltd., Männedorf, Switzerland) instrument in triplicate. The bandwidth was 9 mm and temperature 24 °C.

Determination of flavonoid content in extruded buckwheat products was made with the Liquid Chromatography – Time of Flight Mass Spectrometer (LC–TOF–MS) as described by Klavina et al. (2015). During LC–MS analysis, each sample produces its own base peak chromatogram (BPC). In the positive ionization mode each compound can add a proton and can produce its own [M+H]<sup>+</sup> mass spectra or stay in a positively charged state as M<sup>+</sup> molecule. Each compound has its own chemical formula and molar mass. This chemical formula using *Mass Hunter Qualitative Analyses B.07.00* software can be used for the calculation of [M+H]<sup>+</sup>/[M]<sup>+</sup> which is used for the extraction of compounds from the base peak chromatogram. Both values can be compared and the difference ( $\Delta$ ) between them should not be higher than 0.0030. HRMS experiments ensure accurate mass measurements resulting in the removal of background signals of complex matrix interferences. It is useful for non–targeted or retrospective post–targeted identification of unknown compounds by the processing of raw data obtained in different scan modes, including full scan in defined *m/z* windows. The experimental data were handled using the MassHunter version B07.00 software (Agilent Technologies).

The extruded buckwheat products were analysed in three replications.

#### **Data processing**

The data of research were analysed using mathematical and statistical methods of Microsoft Office Excel 14.0. To determine the significant differences between the mean values were used the analyses of variance (ANOVA), *T*–*test* and *P*–*value* at 0.05.

## **RESULTS AND DISCUSSION**

The total phenolic content (TPC) of ingredients used for taste improvement in extruded buckwheat products is given in Table 2.

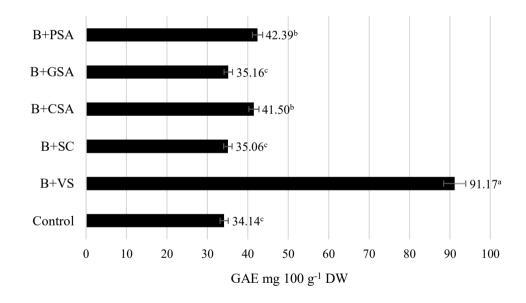
The obtained data allows better to understand and explain the results of current research.

Ingredients for taste improvement	TPC, GAE mg 100 g <sup>-1</sup>		
Sugar	n.d.		
Vanilla sugar	$125.86 \pm 6.09$		
Stevia	n.d.		
Agave syrup	$26.68\pm2.17$		
Cinnamon, g	$1,\!534.07\pm 34.18$		
Caraway	$202.62\pm7.39$		
Garlic powder	$51.60 \pm 1.78$		
Sweet pepper powder	$285.47\pm8.01$		
Salt	n.d.		

Table 2. Total phenolic content in extruded buckwheat products with different taste

n.d. - not detected.

The extruded buckwheat products with sweet and salty taste displayed higher TPC as control sample (Fig. 1). It means that in generally the added ingredients for taste improvement of extruded buckwheat products had favourable effect on TPC. However statistically significant differences were observed between control and extruded buckwheat products with vanilla sugar + stevia; caraway + salt + agave syrup and sweet pepper powder + salt + agave syrup.



**Figure 1.** Total phenolic content in extruded buckwheat products with different taste. Used letters (a, b, c) point to statistically significant difference between buckwheat products (P < 0.05).

The highest TPC (P < 0.05) was determined in extruded buckwheat product with vanilla sugar + stevia (91.17 ± 0.67 GAE mg 100 g<sup>-1</sup> DW). The impressive increase of TPC in extruded buckwheat product with vanilla sugar + stevia could be explained with added vanilla sugar. The results of current research (Table 2) showed, that vanilla sugar had a high total phenolic content (125.86 ± 6.09 GAE mg 100 g<sup>-1</sup>), whereas in stevia it was not detected at all. There are not researches in literature about content of possible phenolic compounds in vanilla sugar. Only Burri et al. (1989) concluded that vanillin

acts as an antioxidant in products of cereals. There are necessary profound researches about vanilla sugar, but it could be concluded, that the addition of vanilla sugar to extruded buckwheat product provided significant increase of TPC.

The great TPC (P < 0.05) in comparison to control was determined in extruded buckwheat products with caraway + salt + agave syrup and sweet pepper powder + salt + agave syrup, too. The increase of TPC in extruded buckwheat products could be based on addition of caraway and sweet pepper powder, because both presented great TPC (202.62 and 285.47 GAE mg 100 g<sup>-1</sup>, respectively). Sharma et al. (2016) reported about high total phenolic content in red sweet pepper, which varied between  $488.81 \pm 8.18$  and  $977.63 \pm 25.40 \ \mu g \ GAE \ g^{-1} FW$  (fresh weight). It is known that production process of sweet pepper powder can markedly affect the total phenolic and flavonoid content, however it is rich source of phenols. Shotorban et al. (2012) indicated that different processing techniques can significantly affect the content of bioactive compounds like polyphenols. Therefore the increase of TPC in extruded buckwheat product with sweet pepper powder + salt + agave syrup was not so marked. The other reason for increase of TPC in extruded buckwheat products with salty taste could be related with addition of agave syrup. Agave syrup is produced from agave plant and it rightly pointed to the presence of TPC in agave syrup (26.68 GAE mg 100 g<sup>-1</sup>). Unfortunately, the few researches in literature about agave syrup chemical composition are focused on sugar composition.

The extruded buckwheat products with added sugar + cinnamon and garlic powder + salt + agave syrup did not provide expected increase of TPC, though cinnamon had the greatest total phenolic content among the added ingredients for taste improvement. It could be explained with the small amount of added cinnamon to dough, and extrusion, which had adverse effects on total phenolic content according to previous research (Beitane et al., 2018).

Extruded buckwheat product with garlic powder + salt + agave syrup had significantly greater concentration of rutin and quercetin (P < 0.05) comparing to other products (Table 3).

	Rutin	Quercetin	Catechin	Epicatechin
Control	$5.51\pm0.72^{\rm b}$	$1.48\pm0.13^{\text{b}}$	$3.97\pm0.23^{\rm a}$	$7.36\pm0.62^{\rm a}$
B+VS	$6.75\pm0.81^{\text{b}}$	$1.68\pm0.12^{\text{b}}$	$4.45\pm0.26^{\rm a}$	$8.02\pm0.69^{\rm a}$
B+SC	$6.72\pm0.77^{\rm b}$	$1.59\pm0.19^{\text{b}}$	$4.01\pm0.20^{\rm a}$	$6.24\pm0.48^{\text{b}}$
B+CSA	$5.35\pm0.90^{b}$	$1.28\pm0.21^{\text{b}}$	$2.20\pm0.18^{\text{b}}$	$4.20\pm0.33^{\circ}$
B+GSA	$8.99\pm0.69^{\rm a}$	$2.27\pm0.17^{\rm a}$	$2.47\pm0.17^{\text{b}}$	$7.26\pm0.52^{\rm a}$
B+PSA	$5.90\pm0.72^{\text{b}}$	$1.52\pm0.11^{\text{b}}$	$2.65\pm0.22^{\text{b}}$	$5.07\pm0.66^{\rm c}$

Table 3. Flavonoids content in extruded buckwheat products with different taste, mg 100 g<sup>-1</sup> DW

Mean values with different letters (a, b, c) within a column point to statistically significant difference between buckwheat products (P < 0.05).

The increase of quercetin concentration in extruded buckwheat product with garlic powder + salt + agave syrup could be based on literature, where Griffiths et al. (2002) reported about high content of flavonoids, especially, quercetin and its conjugates in garlic; Miean & Mohamed (2001) determined relatively high content of quercetin – 47.0 mg kg<sup>-1</sup> DW, myricetin – 693.0 mg kg<sup>-1</sup> DW and apigenin – 217.0 mg kg<sup>-1</sup> DW in garlic. The extruded buckwheat products with garlic powder + salt + agave syrup

contained great concentration of rutin in the present study. It could be due to the presence of added garlic powder and agave syrup, because both presented the content of phenols. Scientific reports about flavonoids composition in garlic did not provide any information about rutin concentration. However the conclusions of researches about flavonoids content in garlic are contrary, for example Beato et al. (2011) did not detect myricetin, quercetin, kaempferol and apigenin in garlic. Therefore it cannot exclude the possibility that the addition of garlic powder provided the increase of rutin content in extruded buckwheat product with garlic powder + salt + agave syrup. The aim of current research was not to evaluate the flavonoid content of added ingredients, therefore it could be investigate in the following research. It was positive that all products except extruded buckwheat product with caraway + salt + agave syrup had rutin and quercetin concentration higher than control sample.

The greatest concentration of catechin and epicatechin was observed in extruded buckwheat product with vanilla sugar + stevia. Product which had the highest TPC, too. Catechin concentration among extruded buckwheat product with vanilla sugar + stevia, control sample and extruded buckwheat product with sugar + cinnamon was insignificant (P > 0.05) as well as epicatechin concentration was insignificant among extruded buckwheat product with vanilla sugar + stevia, buckwheat product with vanilla sugar + stevia.

The extruded buckwheat product with caraway + salt + agave syrup in addition contained luteolin  $-0.24 \text{ mg } 100 \text{ g}^{-1} \text{ DW}$ , kaempferol  $-0.46 \text{ mg } 100 \text{ g}^{-1} \text{ DW}$  and isoquercitrin  $-1.55 \text{ mg } 100 \text{ g}^{-1} \text{ DW}$ . Isoquercitrin was detected in extruded buckwheat products with garlic powder + salt + agave syrup and sweet pepper powder + salt + agave syrup (0.22 and 0.35 mg 100 g^{-1} \text{ DW}, respectively).

The extruded buckwheat product with sugar + cinnamon showed insignificant results about TPC, rutin, quercetin and catechin content comparing to control sample, except epicatechin content, which was significantly lower. The addition of cinnamon did not give the expected result, though Adisakwattana et al. (2011) reported that in cinnamon extract the TPC varied between 0.17–0.21 g GAE g<sup>-1</sup> and flavonoids content – between 48.85–65.52 mg QE g<sup>-1</sup>.

# CONCLUSIONS

The aim to produce new buckwheat product with increased nutritional value was achieved, because some added ingredients used for taste improvement can significantly influence the total phenolic content and flavonoid content. However there is a need for further researches to understand and explain how and why the added ingredients affected/not affected the total phenolic and flavonoid content in extruded buckwheat products.

ACKNOWLEDGEMENTS. This research was supported by National Research Programme AgroBioRes (2014–2017) and the project 'Strengthening Research Capacity in the Latvia University of Agriculture'.

#### REFERENCES

- Adisakwattana, S., Lerdsuwankij, O., Poputtachai, U., Minipun, A. & Suparpprom, C. 2011. Inhibitory activity of cinnamon bark species and their combination effect with acarbose against intestinal  $\alpha$ -glucosidase and pancreatic  $\alpha$ -amylase. *Plant Foods for Human Nutrition* **66**, 143–148.
- Beato, V.M., Orgaz, F., Mansilla, F. & Montano, A. 2011. Changes in phenolic compounds in garlic (*Allium sativum* L.) owing to the cultivar and location of growth. *Plant Foods for Human Nutrition* 66, 218–223.
- Beitane, I., Krumina–Zemture, G & Sabovics, M. 2018. Effect of germination and extrusion on the phenolic content and antioxidant activity of raw buckwheat (*Fagopyrum esculentum* M.). Journal Agronomy Research 16. (Accepted for publication)
- Burri, J., Graf, M., Lambelet, P. & Loliger, J. 1989. Vanillin more than a flavouring agent a potent antioxidant. *Journal of the Science of Food and Agriculture* **48**, 49–56.
- Crawford, P. 2009. Effectiveness of cinnamon for lowering haemoglobin A1C in patients with type 2 diabetes: A randomized, controlled trial. *The Journal of the American Board of Family Medicine* **22**, 507–512.
- Foster–Powell, K., Holt, S.H.A. & Brand–Miller, J.C. 2002. International table of glycemic index and glycemic load values: 2002. *The American Journal of Clinical Nutrition* **76**, 5–56.
- Ghoneem, K.M., Saber, W.I.A., El–Awady, A.A., Rashad, Y.M. & Al–Askar, A.A. 2016. Alternative preservation method against *Sclerotium* tuber rot of Jerusalem artichoke using natural essential oils. *Phytoparasitica* **44**, 341–352.
- Griffiths, G.F., Trueman, C.T., Thomas, B.M. & Smith, B.M. 2002. Onions, a global benefits to health. *Phytotherapy Research* 16, 603–615.
- Herald, T.J., Gadgil, P. & Tilley, M. 2012. High-throughput micro plate assays for screening flavonoid content and DPPH-scavenging activity in sorghum bran and flour. *Journal of the Science of Food and Agriculture* **92**, 2326–2331.
- Hernandez–Ortega, M., Ortiz–Moreno, A., Hernandez–Navarro, M.D., Chamorro–Cevallos, G., Dorantes–Alvarez, L. & Necoechea–Mondragon, H. 2012. Antioxidant, antinociceptive, and anti–inflammatory effects of carotenoids extracted from dried pepper (*Capsicum annuum* L.). *Journal of Biomedicine and Biotechnology*, 1–10. DOI:10.1155/2012/524019
- Hes, M., Dziedzic, K., Gorecka, D., Drozdzynska, A. & Gujska, E. 2014. Effect of boiling in water of barley and buckwheat groats on the antioxidant properties and dietary fiber composition. *Plant Foods for Human Nutrition* 69, 276–282.
- Inglett, G.E., Rose, D.J., Chen, D., Stevenson, D.G. & Biswas, A. 2010. Phenolic content and antioxidant activity of extracts from whole buckwheat (*Fagopyrum esculentum* Moench) with or without microwave irradiation. *Food Chemistry* **119**, 1216–1219.
- Klavina, L., Springe, G., Nikolajeva, V., Martsinkevich, I., Nakurte, I., Dzabijeva, D., Steinberga, I. 2015. Chemical composition of analysis, antimicrobial activity and cytotoxicity screening of moss extracts (Moss phytochemistry). *Molecules* 20(9), 17221– 17243.
- Kluz, M., Terentjeva, M., Puchalski, C., Hutkova, J., Kantor, A., Petrova, J., Mellen, M., Cubon, J., Hascik, P., Kordiaka, R., Kunova, S. & Kacaniova, M. 2016. The extension of shelf-life of chicken meat after application of caraway and anise essential oils and vacuum packaging. *Potravinarstvo* 10(1), 132–138.
- Lin, L.Y., Liu, H.M., Yu, Y.W., Lin, S.D. & Mau, J.L. 2009. Quality and antioxidant property of buckwheat enhanced wheat bread. *Food Chemistry* 112, 987–991.

- Marti, M.C., Camejo, D., Vallejo, F., Romojaro, F., Bacarizo, S., Palma, J.M., Sevilla, F. & Jimenez, A. 2011. Influence of fruit ripening stage and harvest period on the antioxidant content of sweet pepper cultivars. *Plant Foods for Human Nutrition* 66, 416–423.
- Mellado–Mojica, E. & Lopez, M. 2015. Identification, classification, and discrimination of agave syrups from natural sweeteners by infrared spectroscopy and HPAEC–PAD. Food Chemistry 167, 349–357.
- Miean, K.H. & Mohamed, S. 2001. Flovonoid (myricetin, quercetin, kaempferol, luteolin, and apigenin) content of edible tropical plants. *Journal of Agricultural and Food Chemistry* **49**, 3106–3112.
- Mikulajova, A., Sediva, D., Hybenova, E. & Mosovska, S. 2016. Buckwheat cultivars phenolic compounds profiles and antioxidant properties. *Acta Chimica Slovaca* 9, 124–129.
- Muniz-Marquez, D.B., Contreras, J.C., Rodriguez, R., Mussatto, S.I., Wong-Paz, J.E., Teixeira, J.A. & Aguilar, C.N. 2015. Influence of thermal effect on sugars composition of Mexican Agave syrup. *Journal of Food* 13, 607–612.
- Olsson, K., Carlsen, S., Semmler, A., Simon, E., Mikkelsen, M.D. & Moller, B.L. 2016. Microbial production of next-generation stevia sweeteners. *Microbial Cell Factories* 15(207), 1–14. DOI 10.1186/s12934–016-0609–1
- Onyeoziri, U.P., Romanus, E.N. & Onyekachukwu, U.I. 2016. Assessment of antioxidant capacities and phenolic contents of Nigerian cultivars of onions (*Allium cepa* L.) and garlic (*Allium sativum* L.). *Pakistan Journal of Pharmaceutical Sciences* **29**, 1183–1188.
- Otto, A.D. 2010. Cinnamon as a supplemental treatment for impaired glucose tolerance and type 2 diabetes. *Current Diabetes Reports* **10**, 170–172.
- Parkash, D., Suri, S., Upadhyay, G. & Singh, B.N. 2007. Total phenol, antioxidant and free radical scavenging activities of some medicinal plants. *International Journal of Food Sciences and Nutrition* 58, 18–28.
- Phillips, K.M., Carlsen, M.H. & Blomhoff, R. 2009. Total antioxidant content of alternatives to refined sugar. *Journal of the American Dieteric Association* **109**, 64–71.
- Qin, P., Wang, Q., Shan, F., Hou, Z. & Ren, G. 2010. Nutritional composition and flavonoids content of flour from different buckwheat cultivars. *International Journal of Food Science* and Technology 45, 951–958.
- Raybaudi–Massilia, R., Suarez, A.I., Arvelo, F., Zambrano, A., Sojo, F., Calderon– Gabaldon, M.I. & Mosqueda–Melgar, J. 2017. Cytotoxic, antioxidant and antimicrobial properties of red sweet pepper (*Capsicum Annuum* L. var. Llaneron) Extracts: *In vitro* study. *International Journal of Food Studies* 6, 222–231.
- Sensoy, I., Rosen, R.T., Ho, C.T. & Karwe, M.V. 2006. Effect of processing on buckwheat phenolics and antioxidant activity. *Food Chemistry* **99**, 388–393.
- Shannon, M., Rehfeld, A., Frizzell, C., Livingstone, C., McGonagle, C., Skakkebaek, N.E., Wielogorska, E. & Connolly, L. 2016. *In vitro* bioassay investigations of the endocrine disrupting potential of steviol glycosides and their metabolite steviol, components of the natural sweetener *Stevia*. *Molecular and Cellular Endocrinology* **427**, 65–72.
- Sharma, V.K., Chandel, C., Kumar, R., Meena, R.D. & Mawliya, M. 2016. Influence of planting time and fruit maturity stage on antioxidant activity, phenols and anthocyanin contents in sweet pepper. *Research on Crops* 17, 360–368.
- Shotorban, N.I.Y., Jamei, R. & Heidari, R. 2012. Antioxidant activities of two sweet pepper Capsicum annuum L. varieties phenolic extracts and the effects of thermal treatment. *Avicenna Journal of Phytomedicine* **3**, 25–34.
- Unal, H., Izli, G., Izli, N. & Asik, B.B. 2017. Comparison of some physical and chemical characteristics of buckwheat (*Fagopurym esculentum* Moench) grains. *CYTA – Journal of Food* 15, 257–265.

- Vollmannova, A., Margitanova, E., Toth, T., Timoracka, M., Urminska, D., Bojnanska, T. & Cicova, I. 2013. Cultivar influence on total polyphenol and rutin contents and total antioxidant capacity in buckwheat, amaranth, and quinoa seeds. *Czech Journal of Food Sciences* 31, 589–595.
- Wronkowska, M., Honke, J. & Piskula, M.K. 2015. Effect of solid-state fermentation with *Rhizopus oligosporus* on bioactive compounds and antioxidant capacity of raw and roasted buckwheat groats. *Italian Journal of Food Science* 27, 424–431.
- Zabkova, M., Otero, M., Minceva, M., Zabka, M. & Rodrigues A.E. 2006. Separation of synthetic vanillin at different pH onto polymeric adsorbent Sephabeads SP206. *Chemical Engineering and Processing* 45, 598–607.
- Zhang, Z.L., Zhou, M.L., Tang, Y., Li, F.L., Tang, Y.X., Shao, J.R., Xue, W.T. & Wu, Y.M. 2012. Bioactive compounds in functional buckwheat food. *Food Research International* 49, 389–395.
- Zielinska, D., Szawara–Nowak, D. & Zielinski, H. 2007. Comparison of spectrophotometric and electrochemical methods for the evaluation of the antioxidant capacity of buckwheat products after hydrothermal treatment. *Journal of Agricultural and Food Chemistry* **55**, 6124–6131.